EFFECT OF SOME PLANT EXTRACT AGAINST SEED BORNE INFECTION OF COLLECTOTRICHUM DESTRUCTIVUM ON VIGNA UNIGUCULATA L.

Umesh P. Mogle¹ and Sanjay R. Maske²

¹Department of Botany, J. E. S. College, Jalna (M. S.) India ²Department of Botany, K. S. K. College Beed (M. S.) India upmogle@gmail.com

ABSTRACT

The cowpea, *Vigna unguiculata* L. Walp is an ancient food crop, suffering from many fungal diseases. *Collectotrichum destructivum* is a harmful seed borne pathogen causing disease to the cowpea plant. Control of seed borne infection would be a possible means of reducing losses due to this disease, attempts were made, fungal species isolated from cowpea seeds were used as inocula. The effects of leaf extracts of *Argemone mexicana* L., *Semecarpus anacardium* L., *Cassia fistula* L., *Tephrosia purpurea* (L.) Pers., were evaluated for the control of *Collectotrichum destructivum* on seeds of cowpea. The seeds were soaked in sterile distilled water extract (10, 20 and 30%, w/v) of the leaves for 5, 10 and 15 h. All these plant extracts had significant inhibitory growth effect on the fungal pathogen. *Argemone mexicana* extract was more effective followed by *Semecarpus anacardium*, *Cassia fistula* and *Tephrosia purpurea* plant extracts and compared favorably with benomyl in the control of the pathogen.

Key words: Collectotrichum destructivum, Benomyl, Vigna uniguculata.

INTRODUCTION

Cowpea is one of the most ancient human food sources and has probably been used as a crop plant since Neolithic times (Summerfield et al., 1974). It is mainly consumed as a favorite foodstuff in the form of dried seeds, either as flour or split (Johnson and Raymond, 1964; van Wyk and Gericke, 2000). They are a good source of carbohydrates, vitamins, and protein, providing more than half of plant protein in human diets in some areas of the semiarid tropics (Singh et al., 1997; Tuan and Phillips, 1992). The fungus Collectotrichum destructivum attacks all parts of the plant: seedlings, hypocotyl, stems, penduncles, flowers, leaves and pods (Allen et al., 1998). Yield loss of up to 75% due to disease in cowpea has been reported (Emechebe, 1981). Colletotrichum itself was introduced by Corda (1831), It encompasses species with endophytic, epiphytic, saprobic and phytopathogenic lifestyles (Kumar and Hyde, 2004; Photita et al., 2001a,b, 2003, 2004; Liu et al., 2007; Prihastuti et al., 2009). The genus has worldwide importance, causing diseases on a wide range of economic crops and ornamental plants (Sutton, 1992; Than et al., 2008a-c; Hyde et al., 2009). The fungus is seed borne, seed transmitted and causes reduced seed germination (Emechebe,

1981). The control of disease in cowpea has been sought through chemical means and the use of host plant resistance (Oladiran and Oso, 1983; Alabi *et al.*, 1986; Alabi and Emechebe, 1990). However, the average Indian farmer cannot afford the increasing cost of synthetic chemicals. Furthermore, the use of fungicides has of late resulted in the build up of toxic chemicals potentially hazardous to man and environment and also in the build up of resistance by pathogens (Sinclair, 1971; Adesiyan, 1983).

of Therefore, the development biopesticides has been focused as a viable pest control strategy in recent years. One source of potential new pesticides is natural products produced by plants. Plant extracts and essential oils show antifungal activity against a wide range of fungi (Grane & Ahmad, 1988; Wilson et al., 1997; Abd-Alla et al., 2001). Recently Alkhail (2005) showed that aqueous extracts of plants viz., Allium sativum, Cymbopogon proxims, Carum carvi, Azadirachta indica and Eugenia caryophyllus had strong antifungal activity against fungi viz., Fusarium oxysporum, Botrytis cinerea and Rhizoctonia solani. In the present study the antifungal activity of aqueous leaf extracts of four plants against Collectotrichum destructivum was investigated.

MATERIALS AND METHODS

Fresh leaves of Argemone mexicana, Semecarpus anacardium, Cassia fistula, Tephrosia purpurea were collected and washed with distilled water. The solutions were allowed to stand overnight and were strained through a clean muslin cloth. These were centrifuged at 1200 rpm for 20 min. Three concentrations were prepared by extracting 10, 20 and 30 g of plant extracts in 100 ml of sterile distilled water. The seeds, with high infection by C. destructivum as determined by the blotter method, were treated with each of the extracts treatment involving soaking the seeds in each of the concentration for 5, 10 and 15 h. Treated seeds were dried on blotter sheets for 8-10 h and subjected to blotter test. Seeds were also soaked in 3.0% benomyl for 30 min and untreated seeds served as control. For each treatment, four replicates of 25 seeds each were considered making a total of 100 seeds for each treatment. Observation for the incidence of C. destructivum was made under microscope. Cultures and slides were compared with standards. The data on average incidence of C. destructivum were subjected to ANOVA and means were separated using the Duncan's Multiple Range Test (DMRT).

RESULTS AND DISCUSSION

Effect of aqueous leaf extract of four plants viz. Argemone mexicana, Semecarpus anacardium, Cassia fistula, Tephrosia purpurea was evaluated against seed borne infection of Collectotrichum destructivum on Vigna uniquculata L. The three concentrations 10, 20 and 30 of the plant extracts at different time of exposure (soaking hours) reduced the incidence of *C. destructivum*. Complete control was obtained with Argemone mexicana using 30% extracts at 10 and 15h soaking period. There were significant differences in the incidence of C. destructivum on seed treated with the different concentrations of the extracts of S. anacardium and at different time of exposure, soaking periods. The incidence of the pathogen was reduced significantly to 22.0% on cowpea seeds treated with 10% extracts for 15 h. Complete control was achieved by treating cowpea seeds with 20 and 30% extract soaked for 10 and 15 h, respectively (Table 1). Leaf extracts of A. mexicana considerably reduced the fungal incidence to 7.2 and 2.2% in 20% extract after soaking for 10 and 15 h, respectively. The crude extracts from Cassia fistula and Tephrosia purpurea did not completely reduce the incidence of the fungus in all concentrations. However at 30% concentrations, a considerable and significant reduction in the incidence of the pathogen was obtained as 2.0 and 1.0 of C. fistula and 3.0 and 1.5 of T. purpurea. A comparison between the effect of all the plant extracts at 20% concentration (soaking period of 15 h) and benomyl (a standard fungicide), indicated that the extracts from A. mexicana and S. anacardium were superior to the fungicide in achieving disease control in that they completely inhibited the growth of the fungus than the fungicide at 2% concentration and 5. 10 and 15 Min soaking. Most botanical pesticides are known to be general bio-cides or bio-irritants (White, 2004). An in vitro study showed that an aqueous extracts from leaves showed anticancer activity, (Kiranmayi Gali et al., 2011) and antibacterial activity (Kempraj and Bhatt, 2010). Mohanta et al. (2007) prepared the aqueous and organic solvent extracts of the plant S. anacardium and screened for antimicrobial diffusion method) and (disc phytochemical properties. The petroleum ether (PEE) and aqueous extract fractions (AQE) showed inhibitory activity against Staphylococcus aureus (Mona Semalty et al., 2010). C. fistula leaves contents about 72 and 15 % of the anthraquinone glycosides which was 0.62 - 2.01 % ww (Council of Europe, 2000), plant extract contents the laxative potency. All the Cassia fistula extracts were screened for their antibacterial and antifungal activities against the Escherichia coli, Pseudomonas aeruginosa, Staphylococcus aureus, Streptococcus pyogenes and the fungi Candida albicans, Aspergillus niger, and Aspergillus clavatus (Bhalodia and Shukla, 2012). In this study, the plant extracts exhibited anti-fungal activity which varies with varied degrees of concentration and time of exposure to the seeds. Aqueous extracts from leaves of A. Mexicana, S. anacardium and Cassia fistula proved to be more effective in this study against C. destructivum. No work has been reported on the fungitoxicity of this plant extract of T. purpurea on fungus hepatoprotective activity used for human health.

Concentration (%)	Exposure time (h)	Pathogen incidence				
		Benomyl 2%	Argemone mexicana	Semecarpus anacardium	Cassia fistula	Tephrosia purpurea
Control		70.2	70.2	70.2	70.2	70.2
10	5	39.2	30.4	31.4	40.1	41.2
	10	38.1	27.2	26.2	38.1	39.6
	15	35.4	22.2	22.0	37.2	37.2
20	5	17.2	15.4	16.4	19.1	20.2
	10	12.2	7.2	8.5	12.2	12.8
	15	8.2	2.8	4.6	8.4	8.8
30	5	2.5	0.5	2.0	3.8	4.8
	10	1.0	0.0	0.0	2.0	3.0
	15	0.0	0.0	0.0	1.0	1.5

Table 1: Effect of different plant extracts on seed-borne C. destructivum

Values are percentage incidence of *C. destructivum* subjected to square root transformation, and values in the same column followed by the same letters are not significantly different ($p_0.05$ LSD).

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LITERATURE CITED

Abd-Alla, MS, KM Atalla and MAM El-Sawi, 2001. Effect of some plant waste extracts on growth and aflatoxin production by *Aspergillus flavus*. *Annals Agric*. *Sci., Ain Shams Univ., Cairo*, **46**: 579-592.

Alkhail AA, 2005. Antifungal activity of some extracts against some plant pathogenic fungi. *Pak. J. Biol. Sci.,* 8(3): 413-417.

Alabi O, Emecbebe AM and Tyagi Y, 1986. Laboratory and screenhouse evaluation of fungicides for the control of brown blotch of cowpea. *Samaru J. Agric. Rex*, **4**: 25-33

Alabi 0 and Emechebe AM, 1990. Field evaluation of seed treatment and foliar spray fungicides for the control of cowpea brown blotch. *Samaru J. Agric. Res.* **7**: 151-158.

Allen DJ, Thotappilly G, Emechebe AM, Singh BB, 1998. Diseases of Cowpea. In: the pathology of food and pasture legumes. Allen DJ, Lenne JM (ed). Cab international in association with ICRISAT. 750p.

Adesiyan SA, 1983. Pesticides in agro-ecosystem. Proceedings of a Workshop on vertebrate pests and their control in Nigeria (Ed. 0. Funmilayo) pp. 78-90

Bhalodia NR and VJ Shukla, 2011. Antibacterial and antifungal activities from leaf extracts of *Cassia fistula* I.: An ethnomedicinal plant. *J. of Adv. Phar. Tech. and Res.* **2**(2): 104-109.

Corda ACI, 1831. Die Pilze Deutschlands (ed. J. Sturm). Deutschlands Flora, 3. Abtheilung 3: 1-144.

Council of Europe, 2000. *Europeaien pharmacopeia supplement* (3rd Edition). Council of Europe. Strasbourg, pp. 1173-1176.

Emechebe AM, 1981. Brown blotch of cowpea in Northern Nigeria. Samaru J. Agric. Res. 1: 21-26

Grane M and S Ahmad, 1988. *Handbook of plants with pest control properties.* John Wiley and Sons, New York.

Hyde KD, Cai L, Cannon PF, Crouch JA, Crous PW, Damm U, Goodwin PH, Chen H, Johnston PR, Jones EBG, Liu ZY, McKenzie EHC, Moriwaki J, Noireung P, Pennycook SR, Pfenning LH, Prihastuti H, Sato T, Shivas RG, Tan YP, Taylor PWJ, Weir BS, Yang YL and Zhang JZ, 2009. *Colletotrichum* – names in current use. Fungal Diversity **39**: 147-183.

Johnson RM and WD Raymond, 1964. The chemical composition of some tropical food plants II. Pigeon peas and cowpeas. *Tropical Science* 6: 68–73.

Kempraj V and Sumangala KB, 2010. Bacteristatic potential of Argemone mexicana L. against enteropathogenic bacteria. *Indian J. of Natural Products and Resources.* **1**(3):338-341.

Kumar DSS and Hyde KD 2004. Biodiversity and tissue-recurrence of endophytic fungi from *Tripterygium wilfordii*. Fungal Diversity **17**: 69-90.

Kiranmayi Gali, G Ramakrishnan, R Kothai, B Jaykar, 2011. In-vitro Anti-Cancer activity of Methanolic. *Journal of PharmTech Research*, **3**(3):1329-1333.

Liu XY, Duan JX and Xie XM, 2007. *Colletotrichum yunnanense* sp. nov., a new endophytic species from *Buxus* sp. Mycotaxon 100: 137-144

Mohanta TK, Patra JK, Rath SK, Pal DK, Thatoi HN, 2007. Evaluation of antimicrobial activity and phytochemical screening of oils and nuts of *Semecarpus anacardium*. *Sci Res Essay*, **2**:486-90

Oladiran A0 and Oso BA, 1983. Comparative susceptibility of some cowpea tines to brown blotch. *Trop. Grain Legume Bull.* **28:** 10-17

Prihastuti H, Cai L, Chen H, McKenzie EHC and Hyde KD, 2009. Characterization of *Colletotrichum* species associated with coffee berries in Chiang Mai, Thailand. Fungal Diversity **39**: 89-109.

Photita W, Lumyong S, Lumyong P, Ho WH, McKenzie EHC and Hyde KD, 2001a. Fungi on *Musa acuminata* in Hong Kong. Fungal Diversity **6**: 99-106.

Photita W, Lumyong S, Lumyong P and Hyde KD, 2001b. Endophytic fungi of wild banana (*Musa acuminata*) at Doi Suthep Pui National Park, Thailand. Mycological Research **105**: 1508-1514.

Photita W, Lumyong S, Lumyong P, McKenzie EHC and Hyde KD, 2003. Saprobic fungi on dead wild banana. Mycotaxon 85: 345-356.

Photita W, Lumyong S, Lumyong P, McKenzie EHC and Hyde KD, 2004. Are some endophytes of *Musa acuminate* latent pathogens? Fungal Diversity **16**: 131-140.

Sutton BC, 1992. The genus *Glomerella* and its anamorph *Colletotrichum*. In: *Colletotrichum: biology, pathology and control* (eds. J.A. Bailey and M.J. Jeger). CAB International, Wallingford: 1-26.

Mona Semalty, Ajay Semalty, Ashutosh Badola, Geeta Pant Joshi, MSM Rawat, 2010. *Semecarpus anacardium* Linn.: A review. *Pharmacognosy Review*, **4**(7): 88-94.

Sinclair JB, 1971. Fungicides for use on tropical foods. Paper presented at Seminar on plant protection of tropical food crops at University of Tbadan, Ibadan, 29 pp.

Singh BB, DR Mohan Raj, KE Dashiell and LEN Jackai, 1997. Pages x–xii *in* Advances in cowpea research. Copublication of International Institute of Tropical Agriculture (IITA) and Japan International Research Center for Agricultural Sciences (JIRCAS). IITA, Ibadan, Nigeria

Summerfield RJ, Huxley PA, Steele NN, 1974. Cowpea (Vigna unquiculata (L) walp.) Field Crop Abstr. 27: 301-312.

Than PP, Shivas RG, Jeewon R, Pongsupasamit S, Marney TS, Taylor PWJ and Hyde KD, 2008a. Epitypification and phylogeny of *Colletotrichum acutatum* J.H. Simmonds. *Fungal Diversity*, **28**: 97-108.

Than PP, Jeewon R, Hyde KD, Pongsupasamit S, Mongkolporn O and Taylor PWJ, 2008b. Characterization and pathogenicity of *Colletotrichum* species associated with anthracnose on chilli (*Capsicum* spp) in Thailand. *Plant Pathology*, **57**: 562-572.

Than PP, Prihastuti H, Phoulivong S, Taylor PWJ and Hyde KD, 2008c. Chilli anthracnose disease caused by *Colletotrichum* species. *Journal of Zhejiang University: Science* B **9**: 764-778.

Tuan YH and RD Phillips, 1992. Nutritional quality of hard-to-cook and processed cowpea. *Journal of Food Science*, **68**: 1371–1374.

Van Wyk, B-E and N Gericke, 2000. People's plants: a guide to useful plants of southern Africa. Briza Publications, Pretoria, South Africa. 30 pp.

Wilson CL, JM Solar, A El Ghaouth and ME Wisniewski, 1997. Rapid evaluation of plant extracts and essential oils for antifungal activity against *Botrytis cinerea*. *Plant Dis.*, **81**: 201-210.